TOTAL SYNTHESIS OF APRAMYCIN

Kuniaki Tatsuta*, Kohji Akimoto, Hideaki Takahashi, Takao Hamatsu Masahiko Annaka and Mitsuhiro Kinoshita

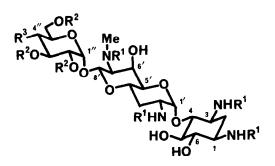
> Department of Applied Chemistry, Keio University Hiyoshi, Kohoku-ku, Yokohama 223, JAPAN

Summary: Apramycin has been stereoselectively synthesized from the previously synthesized aminoglycoside antibiotic, neamine, through the aminooctodiose derivative, aprosamine.

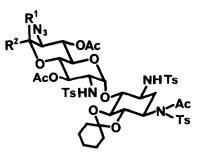
Apramycin (1), ¹ a potent aminoglycoside antibiotic, contains the unusual aminooctodiose moiety. The significant antibiotic activity and unique structure have prompted substantial recent synthesis efforts. 2 We report herein the first total synthesis of apramycin (1) by a route that allows access to a wide variety of structural analogues as well. The synthesis begins with the preparation of the starting masked compound 2^3 (mp 142°, $[\alpha]_D$ -38°) in 79% overall yield from the previously synthesized aminoglycoside antibiotic, neamine 4^{4} , by the following sequence: (1) N-(benzyloxycarbonyloxy)-5-norbornene-2,3-dicarboximide (Z-ONB)⁵/Et₂N/aq. MeOH, 20°, 5h; (2) TsCl/Na₂CO₃/aq. dioxane, 20°, 5h; (3) 1,1-dimethoxycyclohexane⁶/TsOH/DMF, 50°, 25mmHg, 2 days. Saponification of 2 (KO-t-Bu/aq. t-BuOH/70°, 1h) followed by treatment with 30% HCHO and NaBH₃CN⁷ (AcOH/MeCN/20°, 1h) afforded the 6'-N-dimethylamino compound, which after oxidation with m-chloroperbenzoic acid (CHCl₃/20°, 1h) gave the N-oxide (3·H₂O: 83%, mp 179°, $[\alpha]_{D}$ -23°). Treatment of 3 with benzoyl chloride (i-Pr₂EtN/Me₂CO/40°, 10min) produced the aldehyde ($4 \cdot H_2O$: 75%, mp 148°, $[\alpha]_n$ -15°), suggesting that the procedure would be useful for the preparation of aldehydes from the primary amines.⁸ Addition of 15 equiv of allylmagnesium chloride (THF/60°, 0.5h) to 4 unexpectedly 2a,9 provided an approximately 1 : 1 mixture of 6'Sallylcabinol 5 [37%, mp 137°, $[\alpha]_D$ -20°, Rf 0.48 (CHCl₃-MeOH 20 : 1)] and 6'R-isomer 6 (41%, mp 133°, $[\alpha]_{D}$ -5°, Rf 0.43). The stereochemistry at C6' of 5 and 6 was clarified by ¹H-NMR of the derivatives 8a,b and 9 as described below. Oxidation of 5 (OsO₄/aq. dioxane, 20°, 0.5h and then $NaIO_4$, 1.5h) to give the aldehyde [¹H-NMR: 9.79 (CHO)] followed by selective removal of the cyclohexylidene group (50% AcOH in aq. dioxane, 20°, 1.5h) afforded the octodiose 2 derivative $[7 \cdot H_2O: 60\%$, mp 183°, $[\alpha]_D = -13^\circ$ (MeOH)], which was acetylated (Ac₂O/Py, 40°, 16h) to a mixture of tetraacetates 8a [34%, mp 162°, $[\alpha]_D$ +18°, Rf 0.33 (PhH-EtOAc 2 : 1)] and 8b (37%, mp 168°, $[\alpha]_{D}$ +60°, Rf 0.38). Similarly, 6 was transformed into a single β -acetate 9 (mp 169°, $[\alpha]_{D}$ +30°). Their ¹H-NMR decoupling revealed the H-6' methine protons in 8a,b and 9 to be axial $(J_{5',6}^{=10})$ and equatorial $(J_{5',6'}^{=2.5})$, respectively. Their 1-N-acetyl structures were deduced by the H-NMR of the 1-N-acetyl-5,6,3',6',8'-penta-O-acetyl derivative 10 (mp 164°, $[\alpha]_{D}$ +58°), which was formed by removal (50% AcOH in aq. dioxane, 50°, 24h) of the cyclic acetal of 8b followed by acetylation (Ac₂O/Py, 40°, 12h): the significant downfield shift was observed for the H-1 signal (§ 4.34) in comparison with the H-3 signal (§ 3.40). The aforesaid intermediate 7 was converted by our method ¹⁰ (TsCl/Et₃N/MeCN, 20°, 12h and then Ac₂O/Py, 40°, 19h) into the acetyl glycal 11^{11} (52%, mp 164°, $[\alpha]_{D}$ +98°).

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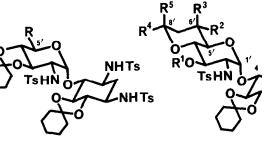
The other isomer 6 was also efficiently transformed into the glycal 11 through the inversion of the 6'-hydroxyl group. Mesylation of 6 (MsCl/Py, 20°, 2.5h) followed by displacement reaction (KOAc/DMSO, 80°, 18h) gave the acetate 12 (85%, mp 138°, $[\alpha]_{p}$ -33°). As described above, oxidation and selective hydrolysis of 12 afforded the octodiose derivative 13 (82%, mp 166°, $[\alpha]_{n}$ +2.5°), which was in turn led to 11 (55%). Azidonitration 12 of 11 [NaN₃/(NH₄)₂Ce(NO₃)₆, -17°, 2.5h, Ar) gave the 7'-azido compounds 14 [63%, mp 161°, [α], -13°, Rf 0.44 (PhH-EtOAc 3 : 1)] and 15 (14%, mp 157°, $[\alpha]_{D}$ +25°, Rf 0.50), the stereochemistry of which was ascertained by their ¹H-NMR. Alkaline treatment of either 14 or 15 in MeOH [Ba(OH)₂·8H₂O, 60°, 25min] afforded the corresponding methyl β -glycoside 16 [mp 160°, [α], +5° (MeOH)] in 71% or 40% yield with the α -anomer. In a straightforward fashion 16 was transformed in 61% overall yield into the 7'-N-(benzyloxycarbonyl)-methylamino derivative 17 (mp 167°, $[\alpha]_{D}$ -44°) via a four-step process : (1) 3 atm H₂/Pt-black/MeOH, 20°, 2h; (2) Z-Cl/Na₂CO₃/aq. Me₂CO, 20°, 10min; (3) LiAlH₄/THF, 80°, 3h; (4) Z-Cl/Na₂CO₃/aq. Me₂CO, 20°, 10min. Mesylation of 17 (MsCl/Py, 8°, 11h; 20°, 5h) afforded quantitatively the labile dimesylate 18 (mp 164°, $[\alpha]_{D}$ +2.5°), which was selectively chlorinated¹³ (LiCl/DMF, 100°, 2h, Ar) to form the 3'-chloro compound, followed by dechlorination (Bu₂SnH/ α , α '-azobisisobutyronitrile/dioxane, 80°, 1.5h, Ar) to give the 3'-deoxy compound 19 (81%, mp 168°, $[\alpha]_{D}$ +13°). Epimerization¹⁴ of the 6'-hydroxyl group was accomplished by treatment of 19 with AcONa (MeOCH_CH_OH, 130°, 2 days) to give the cis-7'-N,6'-O-carbonyl compound 20 (78%, mp 162°, $[\alpha]_{D}$ -25°), the stereochemistry of which was confirmed by the ¹H-NMR ($J_{5',6'}=3.8$, $J_{6',7'}=7$) and IR¹⁴ (1750 cm⁻¹: *cis*-cyclic carbamate). Removal of the Ntosyl group of 20 (Na/liq. NH2, -30°, 1h) followed by alkaline hydrolysis (2M NaOH, 100°, 5h) and subsequent acidic hydrolysis [Dowex 50WX2 (H type) resin/H2O, 20°, 12h and then elution with aq. ammonia] provided the aminooctodiose derivative, methyl $\ddot{\beta}$ -aprosaminide^{la} [21·H₂0: 65%, mp $\sim 145^{\circ}$, [α]_D +109° (H₂O)], which was further hydrolyzed (4N HCl, 95°, 72h) to give aprosamine^{1a} [22·4HCl: 70%, mp 180° (dec.), $[\alpha]_{D}$ +44° (H₂O)]. Both 21 and 22 were identical in all respects with the naturally derived products, thus setting the stage for introduction of the 4-amino-4-deoxy-D-glucose unit. N-benzyloxycarbonylation of 22 (Z-Cl/Na₂CO₂/NaOH/aq.Me₂CO, 20°, 2h) generated the key intermediate [23·H₂O: 71%, mp 195° (dec.), $[\alpha]_D$ +43° (MeOH)]. Methanolysis (1.2% HCl gas in MeOH, 70°, 5h) or methylation (MeI/Ag₂O/MeCN, 20°, 1h) of 23, followed by reduction (3 atm H_2/Pd -black/aq. dioxane, 20°, 5h) gave methyl β -aprosaminide 21 in a fairly good yield (81% or 70%). The findings suggested that the 1,3-diaxial interaction between C6'- and C8'-substituents favored the exclusive formation of the β -glycoside even under acidic conditions.¹⁵ On the other hand, reaction of O-benzylglycosyl halides with alcohols are known to yield predominantly the corresponding α -glycosides.¹⁶ Consequently, it was anticipated that reaction of O-benzylglycosyl halide with 23 would produce the desired α -glycosyl- β -aprosaminide in a reasonable yield. Then, the glycosidation was realized under modified Mukaiyama conditions^{16a} (PhH/dioxane/SnCl₂/AgClO₄/MS 4A, 0°, 2 days) by using 1 equiv of the alcohol 23 and 2 equiv of 4-azido-2,3,6-tri-O-benzyl-4-deoxy-B-D-glucopyranosyl fluoride [oil, $[\alpha]_{D}$ +108°, ¹H-NMR: 5.19 (dd, $J_{1,2}$ =6.4, $J_{1,F}$ =52, H-1)], the latter of which was prepared from the corresponding α -D-glycosyl chloride, ¹⁶ to give the desired glycoside 24 [mp 92°, $[\alpha]_{D}$ +63°, Rf 0.41 (CHCl₃-MeOH 12 : 1); 23: Rf 0.35] in 21% yield (37% yield based on unrecovered alcohol 23. Hydrogenolysis of 24 [3 atm H₂/Pd-black/dioxane-AcOH-H₂O (3 : 1 : 1), 20°, 2h] followed by column chromatography on Amberlite CG-50 (NH $_4$ type) resin with aq. ammonia (0 \rightarrow 0.1M) completed the synthesis, giving apramycin $[1 \cdot H_2 CO_3 \cdot H_2 O: 74\%, mp 240^{\circ} (dec.), [\alpha]_D +162^{\circ}$



1 (Apramycin): R¹ = R² = H, R³ = NH₂ 24: R¹ = Z, R² = BzI, R³ = N₃

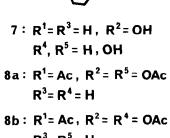


14: $R^1 = ONO_2$, $R^2 = H$ 15: $R^1 = H$, $R^2 = ONO_2$



2: $R = CH_2 NH Z$ 3: $R = CH_2 N(O) Me_2$ 4: R = CHO5: R = H $5^{7'} + H$ 6: R = H

12: R =

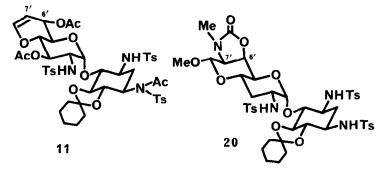


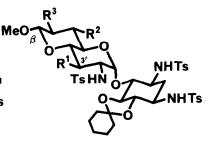
NHTs

$$R^{3} = R^{3} = H$$

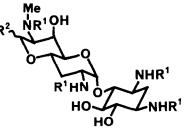
9 : $R^{1} = Ac$, $R^{2} = R^{5} = H$
 $R^{3} = R^{4} = OAc$

13 : $R^1 = R^3 = H$, $R^2 = OAc$ R^4 , $R^5 = H$, OH





16: $R^{1} = R^{2} = OH$, $R^{3} = N_{3}$ 17: $R^{1} = R^{2} = OH$, $R^{3} = N < M_{e}$ 18: $R^{1} = R^{2} = OMs$, $R^{3} = N < M_{e}$ 19: $R^{1} = H$, $R^{2} = OMs$, $R^{3} = N < M_{e}$



21: R¹= H, R²=β-OMe 22 (Aprosamine): R¹= H, R²= OH 23: R¹= Z, R²= OH (H₂O)] which was identical in all respects (TLC, IR, ¹H-NMR and antibacterial activity) with

the authentic sample.

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References and Notes:

- 1) a) S. O'Connor, L. K. T. Lam, N. D. Jones & M. O. Chaney, J. Org. Chem., 41, 2087 (1976). b) R. Q. Thompson & E. A. Presti, Antimicrob. Ag. Chemother. -1967, 332 (1968).
- 2) a) H. C. Jarrell & W. A. Szarek, Can. J. Chem., 57, 924 (1979). b) H. A. Kirst, B. A. Truedell & J. E. Toth, Tetrahedron Lett., 22, 295 (1981).
- 3) All compounds have been fully characterized by spectroscopic means and elemental analyses. Rf-values were measured on silica gel Merck TLC 60F-254.1 Melting points were uncorrected. Optical rotations were done in CHCl₃ at c 1.00 (23°) and H-NMR spectra (250MHz; δ , ppm from TMS, and J in Hz) were in CDCl3 (with or without D₂O) solution, unless stated otherwise. Significant ¹H-NMR spectral data are the following. 1: (5% ND₃-D₂O) 4.29(broad s, 1H, H-6'), 4.92(d, J=8.5, H-8'), 5.15 (d, J=3.5, H-1'), 5.37(d, J=3.5, H-1"). 2: 5.12(s, CH₂Ph), 5.26(d, H-1'). 3: 3.24 & 3.31(s, Me₂N), 5.45(d, J=3, H-1'). 4: 2.41(s, Ts), 9.67(s, CHO). 5: 3.97(dd, J=9.5 & 7, H-5'), 5.11(dd, J=10 & 2, H-9'), 5.14(dd, J=17.5 & 2, H-9'), 5.90(m, H-8'). 6: 4.12(dd, J=10 & 3.5, H-5'), 5.09(dd, J=8 & 1.5, H-9'), 5.14(dd, J=15 & 2, H-9'), 5.88(m, H-8'). 7: (CD₃CN+D₂O) 4.69(dd, J=10 & 2, H-8'ax). 5.08 & 5.09(d, J≃4, H-1'), 5.21 (dull d, J=3, H-8'eq). 8a: 3.48(dt, J=10, 10 & 3.8, H-2'), 3.95(t, J=10, H-5'), 4.53(dt, J= 10.5, 10.5 & 4.5, H-1), 6.07(dull d, H-8'). 8b: 3.83(t, J=10, H-5'), 4.54(dt, J=10.5, 10.5 & 4. H-1). 4.95(m, H-6'). 5.28(d, J=3.5, H-1'), 5.60(dd, J=10 & 2, H-8'). 9: 4.36(dd, J=10 & 2.5, H-5'), 5.59(m, H-6'), 5.85(dd, J=9.5 & 3, H-8'). 10: 3.40(m, H-3), 3.80(t, J=9.5, H-4), 4.34(m, H-1), 5.22(t, J=9.5, H-5), 5.56(dd, J=10.5 & 2.5, H-8'), 5.65(t, J≅10, H-6). 11: 4.84(dd, J=6 & 2.3, H-7'), 5.32(m, H-6'), 6.29(dd, J=6 & 1.5, H-8'). 12: 3.77(dd, J=11 & 8.5, H-5'), 5.07(dd, J=10 & 2, H-9'), 5.12(dd, J=16 & 2, H-9'). 13: 1.98(s, Ac), 2.35 & 2.41 (s. Ts). 14: 3.94(dd, J=10.5 & 4, H-7'), 5.40(t, J≅10, H-6'), 6.20(d, J=4, H-8'). 15: 3.50 (dd, J=10 & 9, H-7'), 5.13(t, J=10, H-6'), 5.43(d, J=9, H-8'). **16**: $[(CD_3)_2CO+D_2O]$ 3.47(t, J=9.5, H-6'), 3.53(s, OMe), 4.21(d, J=8, H-8'), 5.34(d, J=3.5, H-1'). **17**: 3.03(t, J=9.5, H-1). 7'). 3.22(s, NMe). 18: 2.80 & 2.96(s, Ms), 3.00(s, NMe). 19: (50°) 2.76(s, Ms), 3.00(s, NMe). 20: (CD₃CN+D₂O) 3.52(dd, J=7 & 6, H-7'), 4.22(dd, J=10.5 & 3.8, H-5'), 4.42(d, J=6, H-8'), 4.71(dd, J=7 & 3.8, H-6'), 4.93(d, J=3.5, H-1'). 21: $(5\% \text{ ND}_3\text{-D}_2\text{O})$ 2.46(dd, J=8.5 & 3, H-7'), 3.64(dd, J=10 & 2.5, H-5'), 4.26(t, J≅3, H-6'), 4.52(d, J=8.5, H-8'). 22·4HCl: (D_2O) 4.51 & 4.58(t, J=3, H-6'), 5.20(d, J=9, H-8'ax), 5.47(d, J=4, H-8'eq), 5.75(d, J=3, H-8'eq), 5.75(d, J=3, H-8'eq) H-Ĩ') & 5.77(d, J=3.5, H-1'). 23: (CD₃CN+D₂O) 3.00(s, NMe), 4.21(broad s, H-6'). 24: $(CD_3CN+D_2O, 50^\circ)$ 3.73(dt, J=13, 4 & 4, H-2ⁱ), 5.24(d, J=4, H-1ⁱ), 5.27(d, J=9.5, H-8ⁱ), 5.41(d, J=3, H-1").
- 4) K. Tatsuta, E. Kitazawa & S. Umezawa, Bull. Chem. Soc. Jpn., 40, 2371 (1967);
- A. Harayama, T. Tsuchiya & S. Umezawa, Bull. Chem. Soc. Jpn., 52, 3626 (1979).
- 5) T. Naito, S. Nakagawa, Y. Narita & H. Kawaguchi, J. Antibiot., 29, 1286 (1976).
- 6) F. H. Bissett, M. E. Evans & F. W. Parrish, Carbohydr. Res., 5, 184 (1967).
 7) R. F. Borch & A. I. Hassid, J. Org. Chem., <u>37</u>, 1673 (1972).
- 8) The similar strategy was independently reported: K. Takabe, T. Yamada & T. Katagiri, Chem. Lett., 1982, 1987; K. Takabe & T. Yamada, Chem. Ind., 1982, 959. D. A. Evans, J. V. Nelson & T. R. Taber, Top. Stereochem., 13, 1 (1982).
- 10) K. Tatsuta, K. Fujimoto, M. Kinoshita & S. Umezawa, Carbohydr. Res., 54, 85 (1977).
- 11) The glycal 11 was treated with 1% HCl in aq. dioxane (50°, 2 days), followed by acetylation $(Ac_2O/Py, 40^\circ, 12h)$ to give 10 as one of the anomeric acetates, supporting the l-N-acetyl structure.
- 12) R. U. Lemieux & R. M. Ratcliffe, Can. J. Chem., 57, 1244 (1979).
- 13) T. Miyake, Y. Takahashi, T. Tsuchiya, S. Umezawa & H. Umezawa, Nippon Kagaku Kaishi, 1982, 1706.
- 14) H. Sano, T. Tsuchiya, S. Kobayashi, H. Umezawa & S. Umezawa, Bull. Chem. Soc. Jpn., 50, 975 (1977).
- 15) R. U. Lemieux, "Molecular Rearrangement Part II"; P. de Mayo Ed.; Interscience: New York, 1963, p.709.
- 16) a) T. Mukaiyama, Y. Murai & S. Shoda, Chem. Lett., 1981, 431. b) Y. Nishimura, T. Tsuchiya & S. Umezawa, Bull. Chem. Soc. Jpn., 46, 1263 (1973).

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